A comparative study of ectoparasites occurrence between grass carp and silver carp in guilan province culture ponds, Iran

Mehrdad Asgharnia[†] and Mohaddes Ghasemi

Iranian Fisheries Science Research Institute, Inland waters Aquaculture Research Center, Agricultural Research Education and Extension Organization (AREEO), Bandar-e-Anzali, Iran

Parasitic infection is among the most common problems for carp cultivation. They are also important for the principal entrance of other hazardous infections as well. This study was carried out for determining of parasitic fauna of two major carp known as silver and grass carp with the comparison of prevalence value and intensity rate of parasites among them, alongside the relationship between the biometric characteristics and host sex with the infection level. For this purpose, a total of 94 fish samples were caught randomly using a fishing net, from Guilan ponds during spring and summer of the year 2018 and transported alive to the laboratory. Upon arriving, the biometric characteristics and genus of each carp were measured individually. Specimens were then acquired from the skin, gills, and eyes of the carp and examined according to standard parasitology methods. Recovered parasites were observed under a light microscope and then fixed for identification. As the result, the occurrence and intensity of total parasites in male carp were higher than in females. In this research, *Dactylogyrus hypophthalmichthys* and *Dactylogyrus aristhichtys* were observed in silver carp and *Support and Dactylogyrus lamellatus* was detected in grass carp. In the paper below, we found that the host specificity varies in different species of Dactylogyrus isolated from grass carp and silver carp.

Key words: Disease, Parasitic fauna, Host, D. lamellatus, Ctenopharyngodon idella, Cultivation

Introduction

The province of Guilan represents a hub of carp breeding in the country. Most low-income people prefer to eat farmed fish rather than marine or river fish. The herbivorous grass carp (*Ctenopharyngodon idella*) had been introduced into Iran aquaculture for biological control of aquatic weeds harboring the waterways. There are various diseases and infections involving carp that cause mortality and heavy losses in the region. Parasites can have a considerable impact on growth and behavior of fish (Scholz, 1999) and cause mortality in cases of high parasitic burden. This phytophagous fish was noticed to be a target for many parasitic forms (Musselius and Strelkov, 1968; Molnar, 1971; Stepanova,1971; Shirman and Smith, 1983; Abdel-Meguid, 1989, 1995). Dactylo-gyrus has been shown to cause mass mortality of fry, small and brood fish (Paperna, 1991). *Dactylogyrus* spp. are one of the most important parasitic pathogens causing gill-rot disease of grass carp (Bao *et al.*, 2016). Cudmore and Mandrak (2004) stressed that, a large number of disease-causing organisms such as *Dactylogyrus lamellatus* are known to infect Grass carp. It can induce severe damages to the gill filaments by causing hyperplastic proliferation, vacuolar degeneration, edema and necrosis (Molnar,

[†]Corresponding author: Mehrdad Asgharnia

E-mail: mehrdad_asgharnia@yahoo.com

1971; Ramadan *et al.*, 1995). The aim of this paper was to identify the parasitic fauna of two carp species, grass carp and silver carp, by investigating parasite infection on a number of warm water fish ponds in Iran aquaculture, as well as their epizootiological aspects through calculating their prevalence, intensity, abundance, and dominance. In the following manuscript, we discuss host-specific intensity conditions on certain prominent *Dactylogyrus* spp. in a couple of carp species.

Material & methods

Fish samples were caught by random selection, using a fishing net, from Guilan warm water fish ponds during spring and summer of the year 2018 and transported alive to the laboratory. For this purpose, 94 specimens of grass carp and silver carp Hypophthalmychthys molitrix were studied. Upon arriving, the biometric characteristics and sex of each carp were measured individually. Specimens were then acquired from the skin, gills, and eyes of fish and examined carefully according to the standard parasitology method. Recovered parasites were observed through a light microscope and then fixed for identification. Methods used for collecting, fixing, staining, and mounting of parasite specimens were as follows: For the detection of protozoa, the mucus was (scraped separately from the skin, gills) onto a microslide and then spread the mucus with a cover slip. The protozoa to exposed a fixative for about 15 minutes and then washed for several minutes in alcohol containing a drop of added Iodine solution. Next, after dehydration both wet and dry smears were mounted in Canada balsam in accordance with Fernando et al., (1972). Monogenea; fish gills were cut out and examined under a microscope at 40x-100x magnification. Vigorously moving worms were separated from the gills with a pipette and fixed under a coverslip according to Fernando et al., (1972) and Gussev (1983) in ammonium picrate and glycerol-gelatine

respectively. Digenea; metacercaria were collected in a 0.6% saline solution. The sample was placed with a little saline on a glass slide and appropriate pressure applied. It was fixed with 90% alcohol and washed in 70% alcohol and then stained with alum haematoxylin according to Fernando et al., (1972). The identification of whole parasites was achieved in accordance with the keys given by Gussev (1985) and Lom & Dykova (1992). Classical epidemiological variables (prevalence, intensity and abundance) were calculated according to Bush et al., (1997). The length sizes of the all fish were divided into two group ranges; $12.5 \le L_1 \le 36.5$ and $37.5 \le L_2 \le 49$. Results are expressed as mean±standard deviation (SD). Kruskal-Wallis H Test analysis of variance was applied to the data to determine significant differences in the prevalence, intensity and abundance of parasites concerning sex and length groups. Statistical analyses were performed by using the SPSS 22.0 version program and the confidence was kept at 95% level. Typically, a value of less than 0.05 is considered significant.

Results and Discussion

In this study, a total of 94 individual fish (70 silver carp and 24 grass carp) were randomly collected from Guilan warmwater fish ponds. Of these about 40 out of 94 fish were infected with different parasite species. The total number of parasites was 98. The number of infected fish in silver carp was 22 and in grass carp was 18. Dactylogyrus species recovered from silver carp were D. hypophthalmichthys and D. aristhichtys and those in grass carp was D. lamellatus (Table 1). The occurrence, intensity, abundance, and parasite range in the higher length group comparatively were greater than those in comparison with the lower length group (Table 2), but statistically ($\rho=0.439$) not significant (p>0.05). Also, the result declared that the prevalence, intensity, abundance, and parasite range of total parasites in male

Parasite infection	Prevalence value (%)	Mean intensity ± SD	Mean abundance ± SD	Parasite range	Fish species
Diplostomum spathaceum	2.12	1 ± 0	0.02±0.14	1	
Dactylogyrus spp. (hypophthalmichthys	23.4	5.63±4.15	1.31 ± 3.09	1-14	
& aristichthys)					Hypophthalmychthys
Ichthyophthirius multifiliis	4.25	3.5±2.12	0.14 ± 0.77	2-5	molitrix
Trichodina sp.	-	-	-	-	N=70
Lernaea sp.	-	-	-	-	
Nematode	2.12	2 ± 0	0.04 ± 0.29	2	
Diplostomum spathaceum	0.68	2±0	0.04±0.29	2	
Dactylogyrus lamellatus	17.02	9.62 ± 5.50	1.63 ± 4.23	1-16	Ctenopharyngodon
Ichthyophthirius multifiliis	-	-	-	-	idella
Trichodina sp.	2.12	5±0	$0.10{\pm}0.72$	5	N=24
Lernaea sp.	6.38	10.3 ± 4.50	0.65 ± 2.72	6-15	
Total	42.5	12.35±11.81	5.25±9.78	1-34	94

Table 1. Infection amount of parasites in silver carp and grass carp based on different Parasite species from the Guilan province farms in spring and summer of the year 2018

Table 2. Variation of occurrence and infection intensity of parasites in silver carp and grass carp due to average length groups from the Guilan province farms in the spring and summer of the year 2018

Length*	$L_2 = 42.41 \pm 3.78$	$L_1 = 29.37 \pm 7.24$	$L_t = 36.7 \pm 8.53$
Infection	N=42	N=32	*N=74
Occurrence %	14.86	6.75	21.62
Mean intensity SD±	9.45±8.17	5±3.67	8.06±7.26
Mean abundance $SD\!\pm$	2.81±6.14	$0.67{\pm}2.12$	3.48±6.19
Parasite range	1-29	1-10	1-29

*74 samples lenght out of 94 were measured

*(*p*=0.439) not significant (*p*>0.05)

carp were higher than those in females (Table 3). It is usually known that external parasites constitute the largest group of pathogenic organisms in warm water fish (Snieszko and Axelrod, 1971). It seems that various factors are involved in the severity and infection type of these parasites. The most important of them are; water quality, fish density, host diet, body physiology and parasite life cycle. In this study, the prevalence value of parasites in silver carp, *Hypophthalmychthys molitrix*, was higher than grass carp, *Ctenopharyngodon idella*, but intensity rate, mean abundance, and parasite range were lower (Table 1). Probably the reason is *Ctenopharyngodon* *idella* inhabits near the surface layer of the water column where it is less exposed to parastic agents. The grass carp is an herbivore that initially feeds with zoo and phytoplankton and later exclusively on aquatic plants. Apparently, the prevalence value of *Dactylogyrus* sp. in the gills of silver carp was higher than grass carp in the present research, but other parameters were lower (Table 1). On the contrary, previous studies emphasized that the prevalence of *Dactylogyrus* sp. in grass carp was higher than in silver carp.

In the current study, the occurrence value of *Diplostomum spathaceum* in silver carp was higher than that in grass carp (2.12% and 0.68% respec-

sex	Male N=26	Female N=14	Total *N=40
Occurrence %	50	15	65
Mean intensity SD±	13.3±9.65	11±6.5	12.75±8.76
Mean abundance SD±	6±9.25	1.65 ± 4.5	7.65±9.24
Parasite range	2-30	5-18	2-30

Table 3. Variation of occurrence and infection intensity of parasites in silver carp and grass carp due to sex groups in the spring and summer of the year 2018

^{*}40 out of 94 samples sex were determined

tively), but the intensity rate in silver carp (1) was lower than in grass carp (2) (Table 1). In agreement with our research, on a survey in fish farms of Kerman province, the occurrence of this digenea in silver carp (21.1%) was higher than in grass carp (11.9%) too (Ezatkhah, 2014). Unlike grass carp, silver carp inhabits near the soil bed of the pond, where the cercariae of Diplostomum sp. is located, that's becaues silver carp is more sensitive to monogeneans and digenea rather than grass carp. Eye fluke, Diplostomum sp. is known as a blindness factor in Iran's cultivated system and is now widespread throughout Iran (Pazooki et al., 2007). Kiškaroly and Tafro (1983,1988) established the presence of Diplostomum spathaceum in Ctenopharyngodon idella from fish farms in Bosnia and Hercegovina.

Ichthiophthyrius multifliis is the most dangerous and concordant, the most prevalent protozoan of the country's freshwater fish farms which produces heavy damages to the aquaculture industry especially fingerlings (Fig. 1). In the current research, this parasite was only recovered from silver carp, while the other protozoa, *Trichodina* sp. was only observed in grass carp (Table 1). The disease due to *I. multifiliis*, commonly known as *Ichthyophthiriasis*, is widespread and has been reported from different freshwater fish species in Iran (Pazooki *et al.*, 2006; Jalali, 1997; Raissy *et al.*, 2010). Grass carp fingerlings which are kept in the pond or tank with heavy density are susceptible to be infected by *Ichthyophthirius. Ichthyophthirius multifilis* and *Trichodina pediculus* were identified from the gills of silver carp in Bangladesh (Alam *et al.*, 2012). *Ichthyophthirius* sp. with an occurrence rate of 7.1%, density of 5, and mean abundance of 0.36 was also reported from the skin of grass carp in Bangladesh (Akhter *et al.*, 1997). In the following work, the prevalence, and intensity of *Ichthyophthirius multifilis* in the gills of silver carp were 4.2% and 3.5 respectively, which are lower than the previous study with various hosts and different microhabitat.

The absence of *Trichodina* sp. in the fishpond is an indication of water pollution lack and proper pool sanitary management. *Trichodina* sp. with an occurrence rate of 10.4%, density of 4.1, and mean abundance of 0.43 was recorded from the skin of silver carp in Bangladesh (Akhter *et al.*, 1997). Also, *Trichodina* sp. with an occurrence of 8.9%, density of 5, and mean abundance of 0.45 was detected from

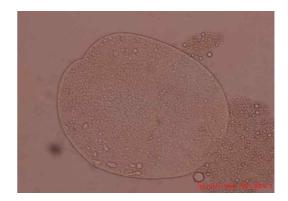


Fig. 1. *Ichthyophthirius multifilis* from the gill of silver carp mag. 100x.

the gill of grass carp in Bangladesh (Akhter *et al.*, 1997). In the available experiment, the prevalence of *Trichodina* sp. (2.1%) in the gills of grass carp was quite lower, but density (5) was consistent with the former inquiry in Bangladesh.

Lernaea species often causes more heavy mortality of bighead (Aristichthys nobilis) rather than grass carp and silver carp, but in this study Lernaea cyprinacea was only separated from grass carp. Yin et al., (1963) found that different species of anchor worms were parasitize on the silver carp and grass carp, as L. ctenopharyngodontis infects grass carp and L. cyprinacea infects silver carp. In the present survey, the prevalence level and intensity of Lernaea sp. in grass carp were 6.3% and 10.3 ± 4.50 respectively. The parasite was isolated from the skin of silver carp in ponds of Bangladesh (Alam et al., 2012). Also, Lernaea sp. was reported from grass carp in Mahabad dam (Abdi et al., 1997). Freshwater fish in Iran are generally encountered by this parasite throughout the year, but severe parasite infestation with mortality occurs only during the warm months of the year and this is due to the fact that ecologically, the evolution of the life cycle of the parasite occurs at high degrees of temperature.

Dove and Ernst (1998) considered that Dactylogyrus is one of the largest genera of parasitic helminths of which 95% are parasites of gills in the fish of the family Cyprinidae. In present study, 3 species of Dactylogyrus spp. were collected from the gills of silver carp and grass carp, consisting D. hypophthalmichthys (Fig. 2), which is host-specific for silver carp, D. aristichthys (Fig. 4) which is host-specific for bighead but separated from silver carp and one host-specific species of D. lamellatus (Fig. 3) recovered from grass carp. The occurrence rate and mean intensity of mixed Dactylogyrus spp. in silver carp were 23.4% and 5.63 respectively, while the occurrence and intensity with Dactylogyrus lamellatus in grass carp were 17.02% and 9.62 respectively. In a study on Kerman province's farms (Ezatkhah,



Fig. 2. *D. hypophthalmichthys* from the gill of silver carp mag. 400x.



Fig. 3. *D. lamellatus* from the gill of grass carp mag. 400x.



Fig. 4. *D. aristichthys* from the gill of silver carp mag. 400x.

2014), the occurrence of *Dactylogyrus* sp. in silver carp (25.3%) was very close to that of the present result, but in the case of grass carp (26.7%) was rela-

tively higher than in our study. In other research on farms of Bangladesh, the occurrence of *Dactylogyrus* sp. in silver carp was 10%, which is lower than the current result, and intensity was 5 which is similar to this work, while the occurrence and intensity of *Dactylogyrus* sp. in grass carp were 13.2% and 4.32 which are lower than our research (Alam *et al.*, 2012). The difference in infection rate in studying fish species may be due to differences in biology, nutrition, the behavior of fish, and also environmental conditions.

D. aristichthys have been reported from fish reproduction centers of the Caspian sea basin (Jalali & Molnar, 1990). D. hypophthalmichthys was identified from silver carp in reproduction centers of Caspian sea basin (Jalali & Molnar, 1990), and D. lamellatus was recovered from grass carp in Behshti hatchery and breeding center (Jalali & Molnar, 1990). Infection of Dactylogyrus lamellatus is responsible for high mortalities observed in fry and fingerling production (Shamsi et al., 2009). Dactylogyrus lamellatus was found in Ctenopharyngodon idella by Sengupta and Dalwani (2008), Shamsi et al., (2009) and Bozorgnia et al., (2012) in fish farms of Iran and by Yao and Nie (2004) in fish farms in China with prevalence higher than 60% and the highest culmination of the mean intensity in April. The highest infection level of D. lamellatus was also observed in a field study in Poland (Pojman'ska, 1995). The host factors like fish size and crowding have a strong influence on infection levels of monogeneans on their fish hosts (Özer and Erdem, 1999). There are different views on the effect of length and weight of the fish on parasitic infection level, some researchers such as our finding (Table 2), believe that infection level increases with increasing weight and length (Bichi and Ibrahim, 2009; Imam and Dewu, 2010), but there is no significant difference between the average length groups and parasite abundance of the examined fish (p>0.05). In the following work, we discovered that host specificity correlation due to silver carp is weaker than against bighead so that, it was able to adopt one species from bighead (*D. aris-tichthys*), in addition to keeping its own host-specific species (*D. hypophthalmichthys*). Conversely, in the case of grass carp, the host-specific severity is so strong that it cannot receive and accept other host-specific species of carp-type Dactylogyrus unless its own specific species, *D. lamellatus*.

Conclusion: In the present research we found that the host specificity varies in different species of Dactylogyrus isolated from grass carp and silver carp. To avoid entry of exotic monogeneans especially *Dactylogyrus* species into the carp fish ponds, introduction of imported live fish to the country should be strictly controlled. Also, to prevent *Dactylogyrusis* and carp infestation with these monogeneans pathogen, the entry of *Carassius gebilio* and goldfish into the carp ponds, due to the fact that *Carassius* species can be the reservoir of many *Dactylogyrus* species for carp species, should be prohibited.

Acknowledgments

The authors of this manuscript wish to express their appreciation for kindness cooperation and financially support throughout the study by both the Iranian National Fisheries Research Sciences Institute and Inland Waters Aquaculture Research Center of Guilan.

References

- Abdel-Meguid, M. (1995). Ectoparasite fauna of grass carp, *Ctenopharyngodon idella* in Delta Breeding Station, Egypt. *Vet. Med. J.* Giza, 43(1): pp. 53-63.
- Abdel-Meguid, M. (1989). Parasitological and histopathological studies on the grass carp, *Ctenopharyngodon idella* raised in the Delta Breeding Station in Egypt. M.s c.Thesis, Ain Shams Univ., Dept. Zoology. 192 p.
- Abdi K., B. Jalali, I. Mobedi and S. Naem. (1997). Survey and identification of crustacean parasites of fish in Mahabad dam. *Pajouhsh va Sazandegi* 3(36):

A comparative study of ectoparasites occurrence between grass carp and silver carp in guilan province culture ponds, Iran 175

pp. 28-32. (In persian)

- Akhter, M. D'Silva, J. and Khatun, A. (1997). Helminth parasites of *Anabas testudineus* (Bloch) in Bangladesh. *Bangladesh J. Zool.* 25: pp. 135-138.
- Alam, M. M., Khan, M. A., Hussain, M. A., Moumita, D., Mazlanand, G. A., and Simon, K. D. (2012). Intensity of parasitic infestation in silver carp, *Hypophthalmichthys molitrix. Journal of Zhejiang University-Science B (Biomedicine & Biotechnology)*, 13(12): pp. 1024-1028.
- Bichi, A. H., & Ibrahim, A. A. (2009). A Survey of Ecto and Intestinal Parasites of Tilapia Zillii (Gervias) In Tiga Lake, Kano, Northern Nigeria. Bayero, *J Pure App Sci*, 2(1): 79-82.
- Bao, J. Y., Hong, Z., Shun, Z., Shan, G. W., Gui, T. W., and Wen, X. L. (2016). Seasonal dynamics and spatial distribution of the *Dactylogyrus* species on the gills of grass carp (*Ctenopharyngodon idellus*) from a fish pond in Wuhan. China, *J. Parasitol*, 102(5): pp. 507-513. DOI: 10.1645/15-931
- Bozorgnia, A. Youssefi, M. R., Barzegar, M. Hosseinifard, S. M., and Ebrahimpour, S. (2012). Biodiversity of Parasites of Fishes in Gheshlagh (Vahdat) Reservoir, Kurdistan Province, Iran. World J. Fish Mar. Sci, 4(3): pp. 249-253.
- Bush, A. O., Lafferty, K. D., Lotz, J. M., and Shostak, W. (1997). Parasitology meets ecology on its own terms: Margolis et al. Revisited. *Journal of Parasitology*, 83, pp. 575-583.
- Cudmore, B., and Mandrak, N. E. (2004). Biological synopsis of grass carp (*Ctenopharyngodon idella*). *Canadian Manuscript Report of Fisheries and Aquatic Sciences* 2705: pp. 1-44.
- Dove, D. M. A., and Ernst, I. (1998). Concurrent invades-faur exotic species of monogenea now established on exotic freshwater fishes in Australia. *Int J Parasitol*, 28: pp. 1755-1764.
- Ezatkhah, M., Alimolaei, M., & Sharifi, H. (2014). A survey of monogenea and digenea trematodes prevalence in warm water cultivated Cyprinidae in Kerman province. *Journal of Aquatic Animals and Fisheries*, 5(18): Summer 2014.
- Fernando, C. H., Furtado, J. I., Gussev, A. V., Hanek, G., and Kakonge, S. A. (1972). "Methods for the Study of Freshwater Fish Parasites", University of Waterloo. Biology Series, Canada. P. 76.
- Gussev, A. V. (1985). Key to parasites of freshwater fishes of the USSR. In: Bauer ON, editor.

Monogenea. Moscow: Nauka Publications, pp. 87-99.

Gussev, A. V. (1983). Methods for collecting and proc-

essing fish parasitic monogenean material. Akad. Nauk. Leningrad, USSR, p. 47.

- Imam, T. S., & Dewu, R. A. (2010). Survey of piscine ecto- and intestinal parasites of clarias species sold at Galadima Road Fish Market, Kano metropolis, Nigeria *Biosci Res Comm*, 22(4): 209-214.
- Jalali, B. (1997). Parasites and parasitic diseases of freshwater fishes of Iran. Iranian fisheries research organization, pp. 105-112. (In Persian)
- Jalali, B., and Molnar, K. (1990). Occurance of Monogenean of Freshwater Fishes of Iran. I. Dactylogyrus from Fish of Natural Waters and Description of Dogielius mokhayeri n.sp. Parasitologica Hungarica, 23: pp. 27-32.
- Kiškaroly, M., and Tafro, A. (1988). Simptomi, patološka slika i dijagnostika najèešæih parazitnih bolesti riba na slatkovodnim ribnjacima. *Vet. Glasnik*, 42(1): pp. 27-33.
- Kiškaroly, M., and Tafro, A. (1983). Ekstenzitet diplostomijaze, najčešće parazitoze riba na ciprinidnim ribnjacima Bosne i Hercegovine. I. *Veterinaria*, 32 (1): pp. 97-103.
- Lom, J., and Dykova, I. (1992). Protozoan Parasites of Fishes (Developments in Aquaculture and Fisheries Science), *Elsevier Science*, Amsterdam, Netherlands, pp. 125-132.
- Molnar, K. (1971). Studies on gill parasitosis of the grass carp (*Ctenopharyngodon idella*) caused by *Dactylogyrus lamellatus* Achmerov, 1952. III. Therapy and control. Acta Veterinaria Academiae Scientiarum Hungaricae, Tomus 21: pp. 361-375.
- Musselius, V. A., and Strelkov, J. A. (1968). Parasites and diseases of the grass and silver carps in fish farms of the U.S.S.R. F.A.O. *Fisheries Report*, 44(5): pp. 353-60.
- Özer, A., and Erdem, O. (1999). The relationship between occurence of ectoparasites, temperature and culture conditions: a comparison of farmed and wild common carp (*Cyprinus carpio* L., 1758) in the Sinop region of northern Turkey. *Journal of Natural History*, 33: pp. 483-491.
- Paperna, I. (1991). Diseases caused by parasites in the aquaculture of warm water fish. Annual Review of Fish Diseases, 1: pp. 155-194.
- Pazooki, J., Masoumian, M., Yahyazadeh, H. M., and Abbasi, K. (2007). Metazoan parasites from freshwater fishes of northeast Iran. *Journal of Agricultural Science and Technology*, 9(1): pp. 25-33.
- Pazooki, J., Masoumian, M., and Jafari, N. (2006). Checklist of Iranian fish parasites. Tehran: *Iranian*

Fisheries Research Organization, (In Persian).

- Pojman'ska, T. (1995). Seasonal dynamics of occurrence and some parasites in four Cyprinid fish cultured in ponds, II. Monogenea. *Acta Parasitologica*, 40(2): pp. 79-84.
- Ramadan, N. F., EL-Banhawy, M. A., and Abdel Salam, M. A. (1995). Parasitological investigation on two monogenetic trematodes and their pathological impact on the grass carp Ctenopharyngodon idella in Egypt. *Journal of the Egyptian German Society of Zoology*, 17: pp. 17-38.
- Raissy, M., Ansari, M., Lashkari, A., and Jalali, B. (2010). Occurrence of parasites in selected fish species in Gandoman Lagoon. *Iran J Fish Sci*, 9(3): pp. 17-21.
- Scholz, T. (1999). Parasites in cultured and feral fish. Veterinary Parasitology, 84: pp. 317-335.
- Sengupta, M., and Dalwani, R. (2008). Identification of worm parasites of fishes in Choghakhor Lagoon in Iran. *The 12th World Lake Conference*, p. 2177-2180.
- Shamsi S, Jalali B, and Aghazadeh Meshgi M. (2009).

Infection with Dactylogyrus spp. among introduced cyprinid fishes and their geographical distribution in Iran. Iran. J. Vet. Res., 10(1): pp. 70-74.

- Shirman, J. V., and Smith, C. R. (1983). Synopsis of biological data on the grass carp, *Ctenopharyngo*don idella. F.A.O. Fisheries Synopsis, No. 135.
- Snieszko, SF., and Axelrod, R. (1971). Diseases of fishes. *TFH Publications*, New Jersey
- Stepanova, G. A. (1971). Parasites of young grass carp, *Ctenopharyngodon idella* at fish farms of the Volga Delta. *Tr. Kasp. Nauchno. Issled Ins. Rybn. Khoz.*, 25: pp. 250-254.
- Yin, W. Y., and *et al.*, (1963). Studies on the Lernaeosis (Lernaea, Copepoda parasitica) of fresh-water fishes of China. *Acta Hydrobiologica Sinica*, 2: pp. 47-117.
- Yao, W. J., and Nie, P. (2004). Population distribution and seasonal alternation of two species of monogeneans on the gills of *Hypophthalmichthys molitrix* and *Ctenopharyngodon idellus*. Acta Hydrobiologica Sinica, 28: pp. 664-667.

Manuscript Received : Aug 15, 2021 Revised : Sep 17, 2021 Accepted : Nov 03, 2021